# 17th INTERNATIONAL SUNFLOWER CONFERENCE

Vol. 1



# Proceedings of the

# 17th International Sunflower Conference

Vol. 1



Córdoba, Spain June 8-12, 2008

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Proceedings of the 17<sup>th</sup> International Sunflower Conference Cordoba, Spain. June 8-12, 2008

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Photography: Spanish landrace of confectionary sunflower collected by L. Velasco and B. Pérez-Vich in Villarta de San Juan, Ciudad Real, Spain, on October 10, 2007.

# Foreword

The proceedings of the 17<sup>th</sup> International Sunflower Conference contain 142 contributions from scientists of 24 countries. They include plenary lectures in several disciplines and regular communications presented in posters during the conference and discussed in the corresponding workshops. The manuscripts are classified by disciplines. They offer a good picture of the current state of the art of sunflower research and cultivation around the world.

The manuscripts in the *Proceedings* have been reviewed by an editorial committee with the main objective of helping the authors to improve their manuscripts through a critical reading. The authors received the edited manuscripts together with the comments of the reviewers and then went on to draft their final version. All the manuscripts received have been published in the *Proceedings*. The contents of the manuscripts are the responsibility of the authors. They should be considered as being privileged communications that require the express consent of the authors to be reprinted in part or as a whole. We wish to thank both the members of the Editorial Committee for their dedication to the task of editing such a large number of manuscripts, as well as all the authors for their collaboration throughout the whole edition process.

The Organizing Committee would also like to thank Diana Badder and José A. Palacios for their excellent editorial assistance in the preparation of these *Proceedings*. We are indebted to the Spanish Association of Sunflower Breeders (Asociación Española de Mejoradores de Girasol), which collaborated actively in the organization of the conference, and, very especially, to Juan Parejo, who was in charge of the financial side.

Finally, we would like to thank all the participants in the conference, who have contributed to its success by a careful preparation and revision of manuscripts and posters, presentation of their research in the workshops, and stimulating discussions throughout the conference on the scientific and technical aspects of sunflower research and cultivation in the world.

The Organizing Committee 17<sup>th</sup> International Sunflower Conference Cordoba, Spain. June 8-12, 2008

# Volume 1 Table of Contents

# PLENARY

Research progress in sunflower diseases and their management  Ferenc Virányi
Could a crop model be useful for improving sunflower crop management?
Francis Flénet, Philippe Debaeke, Pierre Casadebaig
Phenotypic plasticities of yield, phenological development and seed traits Victor Sadras, Abelardo de la Vega.
Sunflower germplasm development utilizing wild <i>Helianthus</i> species Chao-Chien Jan, Gerald J. Seiler, Thomas J. Gulya, Jiuhuan Feng
Current advances in sunflower oil applications Rafael Garcés, Joaquin J. Salas-Liñan, Mónica Venegas-Calerón, Enrique Martínez- Force.
Sunflower in Spain: Past and present trends in an international context  Luis Carlos Alonso
DISEASE RESISTANCE AND PATHOLOGY
Phomopsis control in sunflower using products of biogenic origin  I.I. Begunov, V.T. Piven, A.T. Podvarko, V.Y. Ismailov, T. Gulya
Verticilosis en germoplasma de girasol Julio González, Nora Mancuso, Pedro Ludueña, Antonio Ivancovich
Towards Sclerotinia resistance – In vitro screening of wild sunflower
species
Ksenija T. Ajdukovic, Dragana Miladinovic, Nevena Nagl, Sreten Terzic, Sinisa Jocić, Vladimir Miklič
Correlation between macronutrient content and sunflower resistance to
Sclerotinia sclerotiorum measured by sclerotia infection of stem  Dragana Miladinovic, Slobodanka Pajevic, Ana Marjanovic-Jeromela, Petar Sekulic, Sinisa Jocić, Vladimir Miklič
Races of Plasmopara halstedii on sunflower in separate agrocenosises of
Adigeya Republic, Krasnodar and Rostov regions in Russia Tatiana Antonova, Maria Iwebor, Nina Araslanova.
Differences in some DNA RAPD-loci of <i>Plasmopara halstedii</i> race affecting sunflower in Krasnodar region of Russia  Tatiana Antonova, Saida Guchetl, Maria Iwebor, Tatiana Tchelustnikova
Relations between spring rainfall and infection of sunflower by Plasmopara halstedii (downy mildew)  Denis Tourvieille de Labrouhe, Pascal Walser, Frédéric Serre, Sylvie Roche, Félicits
Vear

	Ea E 9F3	ig aivlig	Giresse,	Xavier	n <b>ch ra</b> elmotte, Pascal <i>N</i>	çois De
among Plasmopar	sqidsnoitsl	netic re	hlight ge	ers hig	mark mebro	erived
				ina Mend	meva, N	eus Du
<i>ານາາດມລາວ</i> ຕິດາ ວວນນ	picical tot	1631 8	1111122 126		uns ui	
nice to Selerotini	staison not	ysnev	когау Бап секоопіп	Madi, IN	ya, bcou	ine skii
				J.	<b>ewoftm</b>	us bəti
k vot resistance i	Uste sinito	r Seler	of enoits	evalua	field	scale
analysis er, Emmanuelle Mestrie uric, François Delmoti	y, Bernard Ga	e Penaud, ire Thiery	he, Annett Serre, Clai	Labrou Tédéric	vieille de Jinard, F	rinoT s pM sən
in sunflower fields A. Leire Molinero-Ru	idtiw nagod	the pat	ersity of	vib bas	enoitil	puos <sub>[1</sub>
opun <i>iipətsim v.v</i>	uomsold 0.	· annetz	riser site	ne virang Te virang	osi, rerei S of th	nozi mi Sangvj
					qnceka	ni əən.
llower triggered b						
і, Сһаѕет Аbbаѕ Аkbат						
activity in sunflow			l shikima Jerotinia			
al tolerance rić, Aleksandra Sudar	ststV sjitsM	t Liović,	<i>inthi</i> Aijié, Ivies	offinus Sis helia A otaA	<i>homop</i> AsinyuC	<b>19/9/17</b> I velsi
					airain	e (Volv
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nbre basal en giras	a boqredun	genética (	stencia g	la resi	eión de	teriza
in sunflower san-Pierre Boudon, Syl	déric Serre, Je	Petit, Fré	s, Aurélie	é Seriey	ar, Herv	city Ve
iccioni, Tomislav Duvnj				Drazenk	randecic,	ıV sailo
ort <i>he helianthi</i> fro	dvia do noi	entificati	cular ide		alicum alicum	
		300 ( <del>570</del> ).	2.5			
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u, Mireille Ducher, De	r, Félicity Vea	eal Walse	ieille, Pasc	viuoT ən	ıkr, Jeanı	Baat Sa
eness in <i>Plasmopa</i> 11, Mireille Ducher, De	s aggressive r, Félicity Vea	est Walser v)? utluence	genes ii V mildev zeille, Paso	<b>01 PU</b> r down	gement nflowe kr, Jean	ganam u <b>s) iib</b> s2 tsed
ат, Denis Tourvieille eness in Plasmopa u, Mireille Ducher, De	aggressive	nfluence v)? sal Walse	i seneg y mildew yeille, Pase	of Pl r down	gement nflowe kr, Jean	ronnor g <b>anam</b> <b>us) iib</b> s2 tasd

Effect of sowing date and initial inoculum of Alternaria helianthi on sunflower in the south region of Brazil
Regina M.V.B.C. Leite, Lilian Amorim, A. Bergamin Filho, Maria Cristina N. de Oliveira, César de Castro.
Effects of nitrogen and water on premature ripening caused by <i>Phoma macdonaldii</i> , a fungal pathogen of sunflower  Célia Seassau, Emmanuelle Mestries, Philippe Debaeke, Grégory Dechamp-Guillaume
Pathological and morphological evaluation of sunflower isohybrids carrying or not the Rcm-1 gene for Sunflower chlorotic mottle virus resistance  Maria Eugenia Bazzalo, Fabián Giolitti, María Teresa Galella, Alberto León, Sergio Lenardon
Study of resistance to <i>Sclerotinia</i> head disease in sunflower genotypes Sándor Páricsi, Melinda Tar, Rozália Nagyné-Kutni
Puccinia helianthi Schw., infecciones en híbridos comerciales en Argentina y su evolución durante dos décadas  N. Huguet, J. Pérez Fernandez, F. Quiroz
Results regarding the influence of <i>in vitro</i> stress induced by the <i>Phomopsis helianthi</i> filtrate on some physiological indices and on sunflower oil quantity and quality
Florentina Raducanu, Elena Petcu, Constantin Raducanu, Maria Stanciu, Danil Stanciu 21
The impact of the new races of broomrape ( <i>Orobanche cumana</i> Wallr.)  parasite in sunflower crop in Romania  Maria Pacureanu Joita, Steluta Raranciuc, Emilia Procopovici, Elisabeta Sava, Dumitru  Nastase
Distribution and dissemination of sunflower broomrape (Orobanche cumana Wallr) race F in Southern Spain  Juan Fernández-Escobar, M. Isabel Rodríguez-Ojeda, Luis Carlos Alonso
ENTOMOLOGY
Development of resistance to insect pests attacking the stem and head of cultivated sunflower in the central and northern production areas of North America  Laurence D. Charlet, Robert M. Aiken, Gerald J. Seiler, Jerry F. Miller, Kathleen A. Grady, Janet J. Knodel
Integrated pest management of the banded sunflower moth in cultivated sunflower in North Dakota  Janet J. Knodel, Laurence D. Charlet
CROP PRODUCTION - MANAGEMENT
Epicuticular wax content in the pericarp of sunflower fruits (Helianthus mnuus L.) grown under moderate water deficit  Maria Clara Franchini, Lilia I. Lindström, Luis F. Hernández 24

••••	V. Thebaud, J.D. Scheiner, J. Dayde
uə	annuus L.) en conditions réelles d'utilisation par les agriculteurs, Midi-Pyrénées - Étude de cas
sni	Impact des facteurs limitants du rendement du Tournesol (Helianth
••••	Silva, Nilza Patricia Ramos
i da	Adriana M. Moreno Pires, Cleide A. Abreu, Aline Renee Coscione, Vinicius Alberti
****	heavy metals
րu	Initial growth of sunflower in soils with high concentrations of boron a
	Walter O. Anyanga, Moses Biruma.
pu	Participatory on-farm sunflower variety evaluation in northern a eastern Uganda
	Edson Perez Guerra.
eira	Maria Carolina Sabbagh, Francine Lorena Cuquel, Ana Cláudia Barneche de Olive Edson Perez Guerra
	əbizonimab
0	Size reduction of ornamental sunflowers by the application
••••	Masahiro Tasumi, Javier Estévez, Pedro Gavilán
.sli <sup>1</sup>	Franco Castillo-Llanque, Cristina Santos, Ignacio J. Lorite, Margarita García-V
HII	of Southern Spain
	Evaluating irrigation performance of sunflower in an irrigation sche
	Insecticide residues cross-contamination of oilseeds during storage Sylvie Dauguet, Jean-Philippe Loison
••	Evandro M. Gomes, Maria Regina G. Ungaro, Dirceu B. Vicira
	-
(T	Sunflower yield and root system development under water stress tropical conditions
	Vladimir Miklič, Siniša Jocić, Dragana Vasić, Nenad Dušanić, Nada Hladni
	affected by harvesting date
B	Changes in seed oil content of sunflower (Helianthus annuus L.)
	J. R. García-Ruiz, F. Perea-Torres, R. Ordóñez-Fernández, J. García-López
	jopo en el girasol
Э.	La agricultura de conservación como sistema viable para combatir
····	Ungaro, Antônio Augusto Lago.
, on	mulch Wilza Patricia Ramos, Maria do Carmo de Salvo Soares Novo, Maria Regina Gonça
u	Sunflower and peanut emergence: initial development under sugarca
	Zvonimir Sakac, Sreten Terzic, Vladimir Miklič
	floral nectar in sunflower (Helianthus annuus L.)
0	The appropriate technique for collecting and measuring the amount
	Branimir Simic, Jasenka Cosic, Ivica Liovic, Miroslav Krizmanic, Jelena Postic
	sunflower hybrids in macro experiments from 1997 to 2007
0	The influence of weather conditions on economic characteristics
••••	Luis F. Hernández
	developed fruits in the sunflower capitulum
(Ia	HAIGHHOOHI HIILM SHOLZOL DAIRAN IA AAHA AAAA AAAA AAAA
g: [ə:	The pattern of foraging paths of the Honey bee (Apis mellifera L.) calse $oldsymbol{A}$

Miyaneh region Ali Faramarzi, Mohammad-Bagher Khorshidi	
Some aspects of sunflower crop management in Romania Gheorghe Sin, Marius Botea, Lenuța Drăgan	
Screening and drying conditions for early harvested sunflower Genta Kanai, Hitoshi Kato, Naonobu Umeda, Kensuke Okada, Morio Matsuzaki	
CROP PRODUCTION – PHYSIOLOGY	
Physiological maturity in sunflower. Correspondence between the quantitative and the visual definition  Luis F. Hernández, Adelina O. Larsen, Lilia I. Lindström, Liliana B. Iriarte	
Influence of desiccation on germination and field emergence of sunflower Ivica Liović, Marijan Bilandžić, Miroslav Krizmanić, Anto Mijić, Ruža Popović, Ilonka Ivanišić, Tomislav Duvnjak, Branimir Šimić, Jasenka Ćosić	
Physiological traits for quantification of drought tolerance in sunflower Elena Petcu, Maria Stanciu, Danil Stanciu, Florentina Raducanu	
El vuelco en el cultivo de girasol: características anatómicas y mecánicas	10
Milena E. Manzur, Antonio J. Hall, Claudio A. Chimenti	
Influence of drought stress on growth, protein expression and osmolyte accumulation in sunflower <i>Helianthus annuus</i> L. c.v. Peredovik Sabine Fulda, Hendrikje Stegmann, Renate Horn	
Development and validation of a model of lodging for sunflower  Mariano M. Sposaro, Pete M. Berry, Mark Sterling, Antonio J. Hall, Claudio A. Chimenti	
Exploring genotypic strategies for sunflower drought resistance by means of a dynamic crop simulation model  Pieme Casadebaig, Philippe Debaeke.	
Dynamics of dry matter accumulation in sunflower Nenad Dusanic, Vladimir Miklič, Igor Balalic, Velimir Radic, Jovan Crnobarac	
IAA/GA3 quantitative ratio of some sunflower genotypes representing	
CMS-Rf system	
Maria Duca, Angela Port.	
Genetic-phytohormonal interactions in male fertility and male sterility phenotype expression in sunflower ( <i>Helianthus annuus</i> L.)  Maria Duca, Angela Port	
Direct and indirect effects of morphophysiological traits on seed yield of	
Nada Hladni, Siniša Jocić, Vladimir Miklič, Anto Mijić, Dejana Saftić Panković	
Determination of maximum achene size in sunflower  A. Mantese, D. Rondanini, D. Medan, A.J. Hall	
Abscisic acid content of a nondormant sunflower (Helianthus annuus L.)	
Brady A. Vick, C.C. Jan	
Producción de girasol ( <i>Helianthus annuus</i> L.) en valles altos de México  L. Alberto Escalante-Estrada, Mª Teresa Rodríguez-González.	
Service Control of the Control of th	

# Correlation between macronutrient content and sunflower resistance to *Sclerotinia* sclerotiorum measured by sclerotia infection of stem

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### **ABSTRACT**

Nutrition of plants has a substantial impact on the predisposition of plants to be attacked or affected by pests and diseases. Since availability and nutrient quantity in the host plants is a limiting factor for *Sclerotinia sclerotiorum* infection development in sunflower, the aim of this work was to determine macronutrient content in photosynthetic tissue (leaves) of sunflower plants before and after development of fungus infection, and to find the correlation between macronutrient content and resistance to sclerotium infection. The study was carried out on eight sunflower inbred lines. Macronutrient (N, P, K, Ca and Mg) content was determined in dry plant material of control and sclerotium infected plants. There was a high positive correlation between N content in infected plants and resistance, which points to the important role of this nutrient in sunflower defence from *Sclerotinia* attack. Moderate negative correlation between K and Ca content in control plants showed that the content of these nutrients in healthy plants could be used as an indicator of cultivated sunflower resistance/susceptibility to *Sclerotinia* infection.

**Key words:** macronutrients – resistance – *Sclerotinia sclerotiorum* – sunflower.

# INTRODUCTION

Nutrition of plants has a substantial impact on the predisposition of plants to be attacked or affected by pests and diseases. By affecting the growth pattern, the anatomy and morphology and particularly the chemical composition, the nutrition of plants may contribute either to an increase or decrease of the resistance and/or tolerance to pests and diseases (Krauss, 2001).

White rot, caused by *Sclerotinia sclerotiorum* (Lib.) de Bary and *S. minor* Jager, is the most important sunflower (*Helianthus amnus* L.) disease in the regions with moderate climate (Masirevic and Gulya, 1992; Vasic et al., 2004). The nutrition of *Sclerotinia* during all stages of disease development is probably the most important factor in determining the extent of infection development (Lumsden, 1979). Smirnova (1967) determined that, for normal development of *Sclerotinia*, all nutrients are needed (N, S, K, Mg, and Fe), while Purdy and Grogan (1954) observed that growth and sclerotium formation only occur when the inorganic macronutrients P, K, Mg and S are present in the medium. Availability and nutrient quantity in the host plants is a limiting factor for *Sclerotinia* infection development in sunflower (Acimovic, 1998).

Because of the importance of nutrients for *S. sclerotiorum* infection development, the aim of this work was to determine the nutrient content in photosynthetic tissue (leaves) of sunflower plants before and after development of fungus infection, and to find the correlation between nutrient content and resistance to sclerotium infection.

# MATERIALS AND METHODS

Six cms inbred lines (PR-ST-3A, CMS<sub>3</sub>-8A, PH-BC<sub>1</sub>-40A, Ha-48A, Ha-74A, and CMS<sub>1</sub>-50A) and two restorer lines (RUS-RF-OL-168 and RHA-583), all selected in Institute of Field and Vegetable Crops, Novi Sad, Serbia, were used in the study.

The experiment was set in two variants -  $SC_1$  - non-infected control, and  $SC_2$  - stem infection with sclerotium. In each variant, four rows with 12 plants per genotype were sown. Plants in  $SC_2$  plot were inoculated by incorporation of sclerotia into the middle part of the stem, at the stage of bud appearance (E4). Wounds with sclerotia were covered with wet cotton and aluminium foil as described by Vasic et al. (2002). After the infection, plants in both variants were irrigated three times a week, for three hours.

Screening was done at the stage of physiological maturity (M0), and resistance was determined as percentage of healthy plants.

Leaf samples for physiological analyses were taken 28 days after the inoculation in both  $SC_1$  and  $SC_2$  variants. In both cases, three healthy leaves from the upper part of the plant were taken from 10 plants from the inner rows. In  $SC_2$  variant, leaves were taken only from the plants showing symptoms of the white rot. Macronutrient (N, P, K, Ca and Mg) content was determined in dry plant material.

Nitrogen content was determined by the Kjehdal method (Nelson and Sommers, 1973) and phosphorus content spectrophotometrically by the ammonium molybdate-vanadate method (Gericke and Kurmies, 1952). Leaf samples were dry-ashed and dissolved in 25% HCl and analyzed for potassium content by flame photometry and for calcium and magnesium content by atomic absorption spectroscopy.

Resistance of genotypes to sclerotia infection was correlated with macronutrient content in infected and non-infected plants in order to estimate their relationship.

### RESULTS AND DISCUSSION

Results obtained showed that there was a difference between tested genotypes in resistance to *Sclerotinia* sclerotium infection. The most resistant genotype was Ha-48A (>90%), and the most susceptible ones were RUS-RF-OL-168 (60.0%) and Ha-74A (60.0%). However, all tested genotypes could be considered tolerant since they all had resistance over 50% (Table 1).

**Table 1.** Macronutrient content in tested sunflower inbred lines.

Genotype	Variant	Resistance	N	P	K	Ca	Mg	
		(%)	mg/100g					
PH-BC <sub>1</sub> -40A	Control		3820	246.7	3223	3069	1250.0	
	Infection	75.0	3582	313.0	3210	3299	1157.0	
Ha-48A	Control		4520	295.7	3478	2873	758.7	
	Infection	93.9	4425	254.3	3112	2883	1043.0	
CMS <sub>3</sub> -8A	Control		3854	288.0	2916	2921	1299.0	
	Infection	77.8	3310	293.7	2969	2572	1265.0	
PR-ST-3A	Control		3691	307.0	3576	2893	866.7	
	Infection	73.7	3834	249.0	2914	2745	1075.0	
CMS <sub>1</sub> -50A	Control		4065	266.7	3646	2796	856.7	
	Infection	60.6	3888	273.7	3516	2623	950.0	
RUS-RF-OL-168	Control		4133	284.0	4552	2991	887.7	
	Infection	60.0	3385	253.0	3181	3694	1397.0	
Ha-74A	Control		3093	323.7	3462	3103	1095.0	
	Infection	60.0	3290	309.0	3620	3646	1271.0	
RHA-583	Control		3412	290.3	4096	2359	619.3	
	Infection	71.9	3643	268.7	3645	2132	580.3	

Nutrient content in sunflower leaf tissue depended both on genotype and the presence or the absence of infection (Table 1). Genotype dependence of nutrient content in sunflower tissue was also observed by other authors (Saric et al., 1991; Vasic et al., 2001).

Nitrogen concentration in leaves of tested genotypes ranged from 4472 mg/100g (average for both variants – infection, control) in genotype Ha-48A, which was at the same time the one most resistant to sclerotium infection (93.9%), to 3192 mg/100g in genotype Ha-74A (Table 1). There was a low positive correlation between resistance and N content in leaves of control plants, and a high positive correlation between resistance and N content in leaves of infected plants (Table 2). This is not in accordance with the conclusions presented in the work of HuiLian (2004). This author connected increased susceptibility of field crop plants to pathogen attack with the increased nitrogen compound content in the plant. However, facultative parasites, such as *Sclerotinia*, require weak plants to infest and kill in order to survive (Marchner, 1995). Vigorous plant growth stimulated by ample N would suppress infestation by this group of pathogens. This may explain the differences in expression of plant diseases in relation to the nutrition of the host and N content.

Phosphorus content in leaves of control plants was in positive correlation with resistance (Table 2). This is in accordance with the results of Sindhan and Parashar (1996), who found that leaves of groundnut (*Apios americana* Medic.) cultivars resistant to *Cercospora arachidicola* contained more phosphorus than leaves of susceptible cultivars.

Potassium concentration in leaves of control plants ranged from 4552 mg/100g in genotype RUS-RF-OL-168 to 2916 mg/100g in genotype CMS<sub>3</sub>-8A (Table 1). Although the results varied, a negative correlation between resistance and K content in leaves of the control, as well as in leaves of infected plants was found (Table 2). Correlation coefficients were moderate and low, respectively, but results clearly showed that a higher K content in leaves has as a consequently lower resistance to sclerotium infection. This is in contrast with the data obtained by Perrenoud (1990) concerning the relationship of K and plant health. This author reviewed some 2450 references and showed that in 70% of all quoted cases K induced a significant reduction in fungal disease incidences. However, in a more recent publication, Mondal et al. (2001) found a negative correlation between K content and disease incidence in soybean (Glycine max (L.) Merrill) and sesame (Sesamum indicum L.).

Table 2. Correlation of resistance of inbred lines to sclerotia infection with macronutrient content in

infected and non-infected sunflower plants.

Macronutrient	Variant	Resistance (%)	Correlation coefficients
N	Control		0.1137
	Infection	75.0	0.5064
P	Control		0.2140
	Infection	93.9	-0.0554
K	Control		-0.3582
	Infection	77.8	-0.1485
Ca	Control		-0.3161
	Infection	73.7	-0.4121
Mg	Control		-0.1917
	Infection	60.6	-0.4040

There was a moderate negative correlation between Ca content in leaves of control and infected plants and resistance to sclerotium infection, meaning that plants with a higher Ca content were more susceptible to the infection (Table 2). This is in contrast with the results obtained by other authors regarding the role of Ca in sunflower resistance and reaction to *Sclerotinia* infection. Antonova et al. (1984) tested chemical composition of leaves and flowers of sunflower plants resistant and susceptible to *Sclerotinia sclerotiorum* and found that flowers of resistant plants contain more Ca.

Infection led to increase in Mg content in leaves of five genotypes (Table 1). Increases in Mg content in infected plants could be a consequence of chlorophyll degradation and disturbed processes of retranslocation and reutilization of Mg ions, since a pathogen attack causes chlorotic to necrotic changes on all plant parts, which are a consequence of chlorophyll degradation (Singh et al., 1998).

Similarly to the results obtained by other authors, nutrient content in leaves of tested sunflower inbred lines was genotype specific. It has also depended on the development of *Sclerotinia* infection, i.e. genotype resistance. There was a high positive correlation between N content in infected plants and resistance, which points to important role of this nutrient in sunflower defence from *Sclerotinia* attack. Moderate negative correlations between K and Ca content in control plants showed that the content of these nutrients in healthy plants could be used as an indicator of cultivated sunflower resistance/susceptibility to *Sclerotinia* infection. Further studies are in progress in order to find out more on the role of nutrients in sunflower response to *Sclerotinia* attack and to test the value of the results of this study on a larger number of sunflower genotypes.

# **ACKNOWLEDGEMENTS**

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## REFERENCES

Acimovic, M. 1998. Sunflower diseases, Institute of Field and Vegetable Crops, Novi Sad, Serbia. (in Serbian).

Antonova, T.S., V.F. Zaichuk, and T.V. Kalinchenko. 1984. Role of calcium in the resistance of sunflower anthodium to grey rot. Selskohozyajstvennaya Biologiya 11:67-68.

- Gericke, P., and S. Kurmies. 1952. Die kolormetriche Phosphorsaürebestimmung mit Ammonium-Vanadate-Molybdat und ihre Anwendung in der Pflanzenanalyse. Z. Pflanzenernähr. Düng. Bodenkd. 59:32-35.
- HuiLian, X. 2004. Accumulation of nitrogen metabolites in relation with incidences of diseases and pest insects in crop plants. p. 1-15. In: S.G Pandalai (ed.), Recent Research Developments in Plant Pathology. International Nature Farming Research Center, Nagano, Japan.
- Krauss, A. 2001. Potassium and biotic stress. p. 281-293. In: Proc 1<sup>st</sup> FAUBA-FERTILIZAR-IPI Workshop, Buenos Aires, Argentina.
- Lumsden, R.D. 1979. Histology and physiology of pathogenesis in plant diseases caused by *Sclerotinia* species. Phytopathology 69:890-896.
- Marchner, H. 1995. Mineral nutrition of higher plants, 2<sup>nd</sup> edn. Academic Press, New York, USA.
- Masirevic, S., and T.J. Gulya. 1992. *Sclerotinia* and *Phomopsis* two devastating sunflower pathogens. Field Crops Res. 30:271-300.
- Mondal, S.S., Pramanik C.K., and J. Das. 2001. Effect of nitrogen and potassium on oil yield, nutrient uptake and soil fertility in soybean (*Glycine max*) sesame (*Sesamum indicum*) intercropping system. Indian J. Agric. Sci. 71:44-46.
- Nelson, D.W., and L.E. Sommers. 1973. Determination of total nitrogen in plant material. Agron. J. 65:109-112.
- Perrenoud, S. 1990. Potassium and plant health. IPI Research Topics No. 3, 2<sup>nd</sup> rev. Edn. Basel, Switzerland.
- Purdy, L.H., and R.G. Grogan. 1954. Physiological studies of *Sclerotinia sclerotiorum* in liquid agar culture. Phytopathology 48:36-38.
- Saric, M., B. Krstic, and D. Škorić. 1991. Element diversity in sunflower inbred lines. Helia 14:41-48.
- Sindhan, G.S., and R.D. Parashar. 1996. Biochemical changes in groundnut leaves due to infection by early and late leaf spot pathogens. Indian J. Mycol. Plant Pathol. 26:210-212.
- Singh, M.J., J. Singh, and S.S. Cheema. 1998. Effect of cucumber mosaic virus on chlorophyll content and mineral elements in chilli. Plant Dis. Res. 13:125-128.
- Smirnova, A.D. 1967. Vlijanije uslovij pitanija na razvitije vozbuditelja sklerotiniji podsolnecnika. Bjulleten Nauc-tehnic informaciji po maslicinim kulturam, p. 10-13.
- Vasic, D., S. Pajevic, M. Saric, Lj. Vasiljevic, and D. Škorić. 2001. Concentration of mineral elements in callus tissue culture of some sunflower inbred lines. J. Plant Nutr. 24:1987-1994.
- Vasic, D., K. Taski, S. Terzic, S. Kevresan, and D. Škorić. 2002. Transferring of *Sclerotinia* resistance from wild into cultivated sunflower combining of conventional and laboratory techniques. Proc. Nat. Sci., Matica Srpska 102:29-33.
- Vasic, D., R. Marinković, F. Miladinovic, S. Jocić, and D. Škorić. 2004. Gene actions affecting sunflower resistance to *Sclerotinia sclerotiorum* measured by sclerotia infections of roots, stems, and capitula. p. 603-608. In: Proc. 16<sup>th</sup> Int. Sunflower Conf., Fargo, ND, USA.