19TH INTERNATIONAL SUNFLOWER CONFERENCE





ISC 2016



PROCEEEDINGS OF 19TH INTERNATIONAL SUNFLOWER CONFERENCE

29 MAY – 3 JUNE, 2016

EDIRNE, TURKEY

19TH INTERNATIONAL SUNFLOWER CONFERENCE

29 MAY – 3 JUNE, 2016, EDIRNE, TURKEY

In Trakya University Balkan Congress Center, Edirne, Turkey

Organized by
Trakya University
and
International Sunflower Association

WELCOME from the CHAIR

You are welcome to our conference that will be jointly organized by Trakya University and International Sunflower Association. The aim of our conference is to present scientific subjects of a broad interest to the sunflower community, by providing an opportunity to present their work as oral or poster presentations that can be of great value for global sunflower production and trade. Our goal is to bring three communities, namely science, research, and private investment together in a friendly environment of Edirne, Turkey in order to share their interests and ideas and to benefit from the interaction with each other.

Our Conference held with record participation with over 600 people working on sunflower as researchers, scientists from seed companies, from oil industry and machinery coming from all part of the World. We have 300 papers which is a record number and almost doubles the previous meetings.

Due to many inquiries about combining our activities with oil industries in ISC 2016, International Sunflower Oil Quality Symposium are organized as one day as a side event during the conference. Sunflower farmers and growers will join also to our conference, so it will be also interesting as an initial attempt to bring together triangle dimensions as scientist, growers and industry in our conference.

Conference activities:

Plenary sessions with oral and poster presentations are on 30^{th} , 31^{st} of May and 1^{st} of June 2016. Besides, the field day and the Sightseeing tours are on June $2^{nd} - 3^{rd}$ June 2016.

Agriculture is an important sector feeding all humankind, but it needs new developments and technologies to supply enough food for increasing world population year by year. Turkey is one of the most important contries on sunflower production and trade and an example to the leading agricultural economies in the world. Therefore, we hope that this conference will help to solve the problems encountered in the Sunflower community with establishing good network collaborations, joint projects and better relationships among countries with sharing our knowledge and experience together. We wish success to this meeting and hope a great scientific achievement together with your contributions.

Edirne is not only a very nice, lovely and historical city at the edge of Europe, but located just at the heart of Balkan region and history endowed with monuments reminding imperial past. We are much pleased to host you all in Edirne and in Turkey.

We would like to thank you to join this conference and we would like to give also special thanks our sponsors and collaborators for giving us big supports to organize this event.

We wish you nice stay in Edirne for truly rewarding days.

Assoc Prof Dr Yalcin KAYA

Head of Organizing Committee
President of International Sunflower Association

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Asst. Prof. Dr. Gokhan KAÇAR	Trakya University	Member
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Dr A. Semsettin TAN	Agean Agric. Res Inst	Member
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Asst. Prof. Dr. Orhan Onur ASKIN	Kirklareli University	Member
Dr Vehbi ESER	BISAB	Member
Kamil YILMAZ	TUBID	Member
Yıldıray GENCER	TURKTOB/TSUAB	Member
Dr Mete KÖMEAĞAÇ	TURKTED	Member
Dr. Maria PACUREANU	Fundulea Agric. Res Inst	Member
Assoc. Prof. Dr. Valentina ENCHEVA	Dobroudja Agric. Res Inst	Member
Dr. Vladimir MIKLIC	Novisad Agric. Res Inst.	Member
Dr. Mehmet DEMIRCI	Agrobest	Member
Mehmet GÜL	Euralis Seed	Member
Ömer IGID	May Seed	Member
Yücel KILIC	Limagrain Seed	Member
Aydın TUNCEL	Pioneer Seed	Member
Abdullah DİŞBUDAK	Soltis Seed	Member
İsmail M. ŞENTÜRK	Syngenta Seed	Member
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Dr. Dragana MILADINOVIC	IFVC Novi-Sad	SERBIA	Weed Management Molecular Breeding
Dr. Siniša JOCIC	IFVC Novi-Sad	SERBIA	Breeding
Dr. Leire MOLINERO-RUIZ	CSIS Cordoba	SPAIN	Disease Resistance
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Dr. Brent HULKE	USDA-ARS Sunflower Research Unit	USA	Breeding
Dr. Lili QI	USDA-ARS Sunflower Research Unit	USA	Molecular Genetics
Dr. Janet KNODEL	North Dakota State Univ.	USA	Sunflower Insects
Dr. Laura MAREK	USDA-ARS Ames, Iowa	USA	Genetic Resources
Dr. Janet KNODEL	North Dakota State Univ.	USA	Sunflower Insects

INVITED SPEAKERS of ISC 2016

SESSIONS

SPEAKER

Breeding Dr Branislav DOZET (Hungary)

Molecular Breeding Dr. Lili QI (USA)

Agronomy and Seed Production Dr Philippe DEBAEKE (France)

Genetic Resources Dr Laura MAREK (USA)

Disease & Pest resistance and Management Prof Dr Steven MASIREVIC (Serbia)

Abiotic Stress Tolerance and Management
Herbicide Resistance and Management
Dr Nicolas LANGLADE (France)
Dr Goran MALIDZA (Serbia)

Confectionery Dr Nada HLADNI (Serbia)

INVITED SPEAKERS of INTERNATIONAL SUNFLOWER OIL QUALITY SYMPOSIUM

NAME	INSTITUTION	COUNTRY
Prof Dr Nurhan T. DUNFORD	Oklahoma State Univ.	USA
Fabrice THURON	Fat & Associes,	FRANCE
Dr Leanordo VELASCO	CSIC, Cordoba,	SPAIN

THE EDITORS OF PROCEEDING BOOK

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Prof. Dr Beraat OZCELIK Istanbul Technical Univ., Turkey

Prof Dr Enrique M. FORCE CSIC, Sevilla, Spain

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Dr Leanordo VELASCO CSIC, Cordoba, Spain

Dr. Yakov DEMURIN Vniimk Institute, Russia

Fabrice TURON Fat & Associes, France

Huseyin BUYUKSAHIN BYSD, Turkey

Metin YURDAGUL MUMSAD, Turkey

Suat OZTURK TYSD, Turkey



19TH INTERNATIONAL SUNFLOWER CONFERENCE 29 MAY – 3 JUNE, 2016 EDIRNE, TURKEY

CONFERENCE PROGRAM

GENERAL SESSION

	SUNDAY, MAY 29 ^m , 2016						
14 ⁰⁰ - 20	Registration at Hotels and Balkan Congress Center						
	MONDAY, MAY 30 th , 2016						
$08^{30} - 09^{30}$	Registration at Balkan Congress Center						
	Opening Ceremony						
10^{30}	Balkan Synphony Orchestra						
	Slide Show: Sunflower from Soil to Table:Our Yellow Bride in the fields						
	Giving Appreciation Certficates to our Sponsors						
$10^{30} - 11$	Coffee break						
00							
11 00 -	OPENING SESSION:						
12 ³⁰	Session Chair: PROF DR MARIA DUCA - Rector of University of Moldova						
	Academy of Science						
11^{-00} –	Invited Speaker Prof Dr. Dragan Skoric "HISTORY OF SUNFLOWER						
11^{40}	BREEDING IN THE WORLD"						
11^{-40} –	Invited Speaker Dr. Lili Qİ "MOLECULAR MAPPING OF THE DISEASE						
12^{20}	RESISTANCE GENES AND ITS IMPACT ON SUNFLOWER BREEDING"						
12^{-20} –	DISCUSSION						
12^{30}							
$12^{30} - 13^{30}$	LUNCH ((Courtesy of Nidera Semillas)						

	GENETIC AND BREEDING	BIOTIC AND ABIOTIC STRESS TOLERANCE	CROP PRODUCTION AND MANAGEMENT	MOLECULAR GENETICS
	(Main Meeting Room)	(2 nd Floor Senate Meeting Room)	(2 nd Floor Left Meeting Room)	(2 nd Floor Right Meeting Room)
	30.05.2016 MONDAY	30.05.2016 MONDAY	30.05.2016 MONDAY	30.05.2016 MONDAY
13 ³⁰ -15 ³⁰	1st Session Chair: CARLOS	1st Session Chair: DR MARIA	1st Session Chair:DR VALENTINA	1st Session Chair: DR RENATE
	FEOLI	JOITA- PACUREANU	ENCHEVA	HORN
13 ³⁰ -13 ⁵⁰	Invited Speaker DR BRANISLAV DOZET Contemporary Challenges in	The genetics and evolution of solar tracking – B. BLACKMAN, S. HARMER	Use of polymer hydrogel in soil moisture conservation for sunflower cultivation in rainfed situations of Northern Karnataka, India: A case study – U. SHANWAD, B. CHITTAPUR, SHANKERGOUD I, B. DESAI, GOVİNDAPPA MR., V. KULKARNI	The cultivated sunflower pan genome provides insights on the wild sources of introgressions and their role in breeding – S. HUBNER, E. ZIGLER, J.R. MANDEL, D. SWANEVELDER, P. VINCOURT, N. LANGLADE, J. M. BURKE, L. H. RIESEBERG
13 ⁵⁰ -14 ¹⁰	Sunflower Breeding	Impact of exogenously applied glycine betaine on physiological attributes of sunflower under drought stress- NOSHIN I., NADIA Z., N. BATOOL, Q. BANO	Determination of the yield and yield components performance of some sunflowers (<i>Helianthus annuus</i> L.) under rainfed conditions – I. DEMIR	Principal Component Analysis for Carbon Isotope Discrimination- Related Traits in Recombinant Inbred Lines of Sunflower – A. L. ADIREDJO, T. LAMAZE, P. GRIEU
14 ¹⁰ -14 ³⁰	M. GHAFFARI	Rapid invitro screening of sunflower genotypes for moisture stress tolerance using PEG 6000 - SHANKERGOUD I., SHESHAIAH K. C.	Appropriate nitrogen (N) and phosphorus (P) fertilizer regime for sunflower (Helianthus annuus L.) in the humid tropics – E. AKPOJOTOR, V. OLOWE	Molecular Studies of Sunflower Responses to Abiotic Stresses – I. TINDAS, R. I. AYTEKIN, S. ÇALIŞKAN
14 ³⁰ -14	Breeding for sunflower hybrids adapted to climate change: the SUNRISE collaborative and multi-disciplinary Project - LUBRANO-LAVADERA A.S., M. COQUE, MUNOS S., DEBAEKE P., MANGIN B., GOUZY J., KEPHALİACOS C., PIQUEMAL J., PINOCHET X.,	Exploring drought tolerance related traits in <i>Helianthus argophyllus</i> , <i>Helianthus annuus</i> and their hybrids – M. MUBASHAR HUSSAIN, M. KAUSAR, M. KHAN, P. MONNEVEUX	Interactive Effects of Different Intra-Row spacing and Nitrogen Levels on Yield and Yield Compenents of confectionery sunflower (<i>Helianthus annuus</i> L.) genotype (Alaca) Under Ankara conditions – S. DAY, O. KOLSARICI	Comparative assessment of androgenic response in sunflower (<i>Helianthus annus</i>) – N. AKGUL, E. ÇABUK ŞAHIN, Y. AYDIN, A. ALTINKUT UNCUOGLU, G. EVCI, A GÜREL

	LANGLADE N.			
14 ⁵⁰ -15 ⁰⁰	Discussion	Discussion	Discussion	Discussion
15 ⁰⁰ -15 ³⁰	Coffee break	Coffee break	Coffee break	Coffee break
15 ³⁰ -17 ⁰⁰	2 nd Session: Chair: DR VLADIMIR MIKLIC	2 nd Session: Chair: DR FELICITY VEAR	2 nd Session Chair: PROF DR GIAN PAOLO VANNOZZI	2 nd Session Chair: DR PHILIPPE DEBAEKE
15 ³⁰ -15 ⁵⁰	Assessment of sunflower germplasm selected for cold tolerance under autumn planting conditions in Morocco - HOUMANAT K., MAZOUZ H., EL FECHTALI M., NABLOUSSI A.	Invited Speaker PROF DR STEVAN MAŠIREVIĆ	Global change adaptation: what future for sunflower crops and products? A foresight study for oilseed chains at 2030 horizon – E. PILORGE, A. M. TREMBLAY, F. MUEL	Molecular and genetic aspects of sunflower defensive response to downy mildew - T. ŞESTACOVA, A.PORT, M. DUCA
15 ⁵⁰ -16 ¹⁰	Perspective and challenges to develop high yielding, disease resistant and oil quality sunflower hybrids in India - R.K.SHEORAN	Sunflower diseases research progress and management	Bioactivity and Phytochemical Evaluation of Sunflower (Helianthus annuus L.) Leaf Extract – Y. BIBI, A. QAYYUM, S. NISA	Recent Molecular Studies on Downy Mildew Disease – A. K. TURKMEN, S. CALISKAN
16 ¹⁰ -16 ³⁰	Stability performance of new introduced sunflower hybrids for seed yield and its components under Sudan conditions – A. A. M. ABDALLA	Control of Verticillium dahliae causing sunflower wilt using Brassica green manures - DESSERRE D., MESTRIES E., DECHAMP-GUILLAUME G., SEASSAU C.	Effects of Different Organomineral and Inorganic Compound Fertilizers on Seed Yield and Some Yield Components of Sunflower (<i>H. annuus</i> L.) – S. SUZER, E. CULHACI	Molecular Studies involved in sunflower responses in drought stres - I. ALTINDAS, E. AKSOY, S. CALISKAN
16 ³⁰ 16 ⁴⁵	Discussion	Discussion	Discussion	Discussion
16 ⁴⁵ -18 ⁰⁰	Poster Session	Poster Session	Poster Session	Poster Session
19 30 -	Dinner Party (Courtesy of Syngenta)	Dinner Party (Courtesy of Syngenta)	Dinner Party (Courtesy of Syngenta)	Dinner Party (Courtesy of Syngenta)

	31.05.2016 TUESDAY	31.05.2016 TUESDAY	31.05.2016 TUESDAY	31.05.2016 TUESDAY
09 ³⁰ -10	3 RD Session Chair: DR OLIVIER COTTET	3 RD Session Chair: PROF DR STEVAN MASIREVIC	3 RD Session Chair: DR AMELIA BERTERO DE ROMANO	3 RD Session Chair: DR DRAGANA MILADINOVIC
09 ³⁰ -09	Collection of wild <i>Helianthus</i> anomalus and deserticola sunflower from the desert southwest USA – G. SEILER, L. MAREK	Isolation and identification of pathogen of Sunflower <i>Fusarium</i> Wilt - JING G. YUAN YUAN Z., GUİ Z., JİAN Z., KAİ W., JUN Z.	Invited Speaker	Proteomic response of sunflower to drought stres – M. GHAFFARI, M. TOORCHI, M. VALIZADEH
09 ⁵⁰ -10	The b1 locus that controls apical shoot branching in <i>H. annuus</i> exhibits a molecular diversity linked to the breeding history of hybrids - DURIEZ P., BONIFACE, M. C., POUILLY N., VAUTRIN S., MAYJ., RODDE N., BERGES H., CARRERE S., GOUZY J., P. VINCOURT, J. PIQUEMAL, S. MUNOS	Distribution of <i>Plasmopara halstedii</i> pathotypes in Hungary – R. BÁN, A. KOVÁCS, G. BAGLYAS, M. PERCZEL, G. TUROCZI, K. KOROSI	Sunflower and climate change in Europe: crop vulnerability, adaptation, and mitigation potential	Identification of HaDELLA, HaGID1 as well as HaSLEEPY and HaSNEEZY genes involved in gibberellin signaling in sunflower - R. EWALD, N. GEHM, L. POPIOLKOWSKI, A. ANTELMANN, R. HORN
10 ¹⁰ -10	Phenotypic and genotypic characterization of 400 new sunflower pre-bred lines – G. BAUTE, W. ANYANGA, E. ALBRECHT, L. H. RIESEBERG	Exploitation of the knowledge on oomycete effectors to drive the discovery of durable disease resistance to downy mildew in sunflower – Y. PECRIX, L. BUENDIA, Q. GASCUEL, C. PENOUILH-SUZETTE, L. GODIARD	Chemical Broomrape (Orobanche cumana) control in Clearfield® sunflower with different Imazamox containing herbicide formulations – M. PFENNING, M. VALTIN, S. SASCHA, J. BESSAI	Characterization of sunflower inbred lines with high oleic acid content by DNA markers – B. B. BILGEN
10 ³⁰ -10	Developing well adapted hybrids in Europe by using a G*E approach - GAUTIER F., HELOISE H., MILAGROS G., SAUVAIRE D.	Response to sunflower (<i>Helianthus annuus</i> L.) plant at early growth stage to cadmium toxicity – Y. CIKILI, H. SAMET, N. C. ATIKMEN	Pulsar® Plus and Eurolightning® Plus - herbicides for enhanced weed control in Clearfield® Plus sunflower – J. BESSAİ, SCHLÄFER S., PFENNING M., MORAN D., CARTIN J.	Evaluation of WRKY and MYB transcription factors in some downy mildew infected sunflower lines; microarray data analysis – E. FILIZ, I. I. ÖZYİĞİT, R. VATANSEVER

$10^{50} - 11^{00}$	Discussion	Discussion	Discussion	Discussion
$11^{00} - 11^{20}$	Coffee break	Coffee break	Coffee break	Coffee break
11 ²⁰ -12 ³⁰	4 th Session: Chair: DR SINISA JOCIC	4th Session Chair: DR MICHAEL FOLEY	4th Session Chair: DR SUJATHA MULPURI	4 th Session Chair: PROF DR RISHI BEHL
11 ²⁰ -11 ⁴⁰	Correlation studies between SSR marker based genetic distance and heterosis in sunflower (Helianthus annuus L.) – V. KULKARNI, SHANKERGOUD I., SUPRIYA S.M, SURESHA P.G.	PCR combined with GFP tagged Verticillium dahliae confirmed the seeds transmission of Sunflower Verticillium Wilt - YUAN YUAN Z., GUI Z., JIAN Z., JUN Z.	Relationships between Germination and Vigor Tests with Field Emergence of Sunflower in Iran – H. SADEGHI, S. SHEIDAEI	Invited Speaker DR STEPHANE MUNOS De novo sequencing of the
11 ⁴⁰ -12 ⁰⁰	Optimization of Agrobacterium- mediated gene transfer systems in Turkish sunflower (<i>Helianthus</i> <i>annuus</i> L.) varieties – I. I. ÖZYİĞİT, S. KARADENIZ, H. TOMBULOGLU, E. FILIZ	Stability of the level of partial resistance to white rot in sunflower – M. ANABELLA DINON, F. CASTAÑO, S. SAN MARTINO, J. LÚQUEZ, F. QUIROZ	Pest Monitoring and Handling System Based on 4G Mobile System – C. ATLIG	Helianthus annuus and Orobanche cumana genomes
12 ⁰⁰ -12 ²⁰	Inclusion of dominance effect in genomic selection model to improve predictive ability for sunflower hybrid performance – F. BONNAFOUS, N. LANGLADE, B. MANGIN	Genetic divergence among sunflower inbred lines and their convergent improvement for yield, quality and disease resistance- R. RANI - R. K. SHEORAN - S. CHANDER - R. K. BEHL	New seed treatment solutions for Plasmospora Resistance Management in Sunflower – F. BRANDL	Comparison of cytoplasmic male sterility based on PET1 and PET2 cytoplasm in sunflower (Helianthus annuus L.) - HORN R., REDDEMANN A., DRUMEVA M
12^{20} - 12^{30}	Discussion	Discussion	Discussion	Discussion
13 ³⁰ -13 ³⁰	Farmer Union)	Lunch (Courtesy of Edirne Farmer Union)	Lunch (Courtesy of Edirne Farmer Union)	Lunch(Courtesy of Edirne Farmer Union)
13 ³⁰ -15 ³⁰	5 th Session Chair: DR THIERRY ANDRE	5 th Session Chair: DR ROBERT NEMETH	5 th Session Chair: PROF DR BENJAMIN BLACKMAN	5 th Session Chair: PROF DR DEJANA PANKOVIC
13 ³⁰ -13 ⁵⁰	Invited Speaker DR MARIA JOITA- PACUREANU Broomrape (Orobanche cumana	Cadmium-potassium interrelationships in sunflower (<i>Helianthus annuus</i> L.) – H. SAMET, Y. CIKILI, N. C. ATIKMEN	Performance of sunflower hybrids in black cotton soils of Northern Karnataka, India – U. SHANWAD, SHANKERGOUD I, S. N. SUDHAKARBABU, V. KULKARNI, GOVINDAPPA	Approaches for improvement of resistance to powdery mildew in sunflower (<i>Helianthus annuus</i> L.) – S. MULPURI, K. PALCHAMY, C. R. SANKARANENI, V.
	Wallr.) - Update on racial		MR, VIJAYKUMAR G.	KODEBOYİNA

13 ⁵⁰ -14	composition and distribution, host resistance and management	Effects of Micro Nutrients (Fe, Zn, B and Mn) on Yield and Yield Components of Two Sunflower (<i>Helianthus annuus</i> L.) Cultivars in Urmia Condition – A. RAHIMI, J. JALILIAN	Modeling sunflower fungal complex to help design integrated pest management strategies - AUBERTOT J. N., MESTRIES E., M. A. VEDY-ZECCHINI, P. DEBAEKE	Genetic engineering studies on sunflower- M. E. ÇALIŞKAN, S. DAS DANGOL
14 ¹⁰ -14	Testing annual wild sunflower species for resistance to <i>Orobanche cumana</i> Wallr – S. TERZIĆ, B. DEDIĆ, J. ATLAGIĆ, S. JOCIĆ, D. MILADINOVIĆ, M. JOCKOVIĆ	Quantification of drought tolerance levels of sunflower inbred lines by means of <i>chlorophyll</i> -a fluorescence - A. S. BALKAN, NALCAIYI, SCULHA ERDAL - O. GUNDUZ, V. PEKCAN, O. ARSLAN, N. CICEK, Y. KAYA, Y. EKMEKCI	Escape to tiny bug (<i>Nysius simulans</i> Stål) attack across planting date adjustment in sunflower hybrid seed crops from southern BuenosAires province, Argentine – J. RENZI, O. REINOSO, M. BRUNA, M. AVALOS, M. CANTAMUTTO	Invited Speaker DR NICOLAS LANGLADE Genome-wide association of oil
14 ³⁰ -14 50	Determination of superior hybrid combinations in sunflower and testing of their resistance to broomrape (<i>Orobanche cumana</i> Wallr.) In infested areas – O. GÜNDÜZ, A. T. GOKSOY	The effect of climate factors and climate change on the yield of sunflower (Helianthus annuus L.) in Marmara region – H. GURKAN, H. BULUT, N. BAYRAKTAR, M. DEMIRCAN, O. ESKİOĞLU, N. KOÇAK	Current Situation, Problems and Solutions of Sunflower in the Central Anatolian Region – C. YAVUZ, S. CALISKAN	yield plasticity to drought, nitrogen and chilling stresses in sunflower
14 ⁵⁰ -15	Discussion	Discussion	Discussion	Discussion
15 ⁰⁰ -15	Coffee break	Coffee break	Coffee break	Coffee break
15 ³⁰ -17	6 th Session Chair: DR CHAO CHIEN JAN	6 th Session: Chair: DR GERALD SEILER	6 th Session Chair: PROF DR MICHELLE GILLEY	6 th Session Chair: DR STEPHANE MUNOS
15 ³⁰ -15	Invited Speaker DR GORAN MALIDZA	Effects of Naphthalene Acetic Acid and N6-Benzyladenine on Androgenesis in <i>Helianthus annuus</i> L. Anthers - S. DAYAN, H. ARDA	Microbial Dressing of Sunflower Seeds with <i>Trichoderma</i> harzianum KUEN 1585 – Y. S. YONSEL, M. SEVİM	QTL mapping for broomrape (<i>Orobanche cumana</i> Wallr.) resistance in sunflower – I. CELİK, D. ZARARSIZ, A. FRARY, S. DOGANLAR
15 ⁵⁰ -16	Integrated weed management in sunflower: Challenges and opportunities	Do cell wall proteins affect the setting of grains and their potential weight in sunflower? – D. CALDERINI, S. VASQUEZ, F. CASTILLO, P.	Green and brown bridges aid survival of multiple <i>Diaporthe / Phomopsis</i> species with a range of virulences on sunflower, soybeans,	Determination the genetic characterization of different lines of sunflower (<i>Helianthus annuus</i> 1.)by using genetic resources

		LIZANA, R. RIEGEL		mungbeans and other Australia. – S. THOM NEATE, Y. PEI TAN SHIVAS, E. AITKEI	IPSON, S.	based on SSRs (Simple Sequence Repeat) – D. BASALMA, M. PASHAZADEH
	Advancements in Clearfield® Plus Sunflower Hybrid Variety Development – B. WESTON, M. PFENNING, C. NIETO, P. ANGELETTI, E. SAKIMA	The Estimating Drought Stress Tolerances of Sunflower Inbred lines under controlled environmental conditions – O. ARSLAN, A. S. BALKAN NALCAIYI, G. EVCI, V. PEKCAN, I. M. YILMAZ, S. ÇULHA ERDAL, N. CICEK, Y. KAYA, Y. EKMEKCI		Evaluation of Sunflow (Helianthus annuus L for Photothermal Unit Accumulation, Oil Yi Quality and Yield Tra Spring Planting Cond Haripur, Pakistan – A I. SULTAN, S. U. KI BIBI, A. MEHMOOL M. A. JENKS	.) Hybrids its eld, Oil its under itions of . QAYYUM, HAN, Y.	Study of the genomic diversity of <i>Verticillium sp.</i> capable of colonizing sunflower. How knowledge of pathogen genetic structure can be combined with classical breeding approaches to guide it – H. MISSONNIER, F. LUIGI, L. GWENAELLE, DAYDÉ J, J. ALBAN, THOMMA B. PHJ
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19 ³⁰ -	Dinner Party		Dinner Party	Dinner Pa	arty Dinner Party	
	01. 06.2016 WEDNESI	DAY	01. 06.2016 WEI	NESDAY	01. 06.2016 WEDNESDAY	
09 ³⁰ -11 ⁰⁰	7 th Session Chair: DR MIGUEL CANTAMUTTO	REGISTRA		TION		
09 ³⁰ -09 ⁵⁰	The effects of applied herbicides of oil quality components of two oled linoleic sunflower (<i>Helianthus an</i> hybrids – F. ONEMLI, U. TETIK	ic and two nuus L.)	INTERNATIONAL SU QUALITY SYMPOSIUM			
09 50-1010	New virulences of <i>Orobanche cur</i> in Romania - PARVU N., TEODO		Session Chair: PROF DR MEHMET EMIN CALISKAN Invited Speaker Fabrice THURON - "HO Oilseeds and Oils Market: Positioning Sunflower Today and Tomorrow			
10 ¹⁰ -10 ³⁰	Genetic characterization of the int between sunflower and <i>Orobanch</i>		Invited Speaker Prof Dr Nurhan TURGUT	DUNFORD		

	PÈREZ-VICH, MUNOS S.		
10 ³⁰ -10 ⁵⁰	Study of <i>Orobanche cumana</i> genetic diversity – M. COQUE, T. ANDRE, R. GIMENEZ, M. ARCHIPIANO, L. POLOVYNKO, M. C. TARDIN, C. JESTIN, B. GREZES-BESSET	Invited Speaker DR. LEONARDO VELASCO Source and sink affect phytosterol concentration and composition of sunflower oil	
10^{50} -11 00	Discussion	Discussion	Discussion
$11^{00} - 11^{20}$	Coffee break	Coffee break	Coffee break
11^{20} - 12^{30}	8 th Session: Chair: DR LOREN H. RIESEBERG	8 th Session: Chair: DR LEONARDO VELASCO	8 th Session: Chair: PROF DR ZHAO JUN
11 ²⁰ -11 ⁴⁰	Invited Speaker	Oil content and oil quality characteristics of linoleic and high-oleic sunflower varieties cultivated in Turkey – B. ASKIN, M. AFACAN, V. BİCER, Ö. KARADAS, İ. KONUK	Quality characteristics of roasted sunflower seeds during storage - M. B. BAHAR, F. SEYHAN, B. OZTURK, B. TOPAL, F. S. BAYRAKTAR
11 ⁴⁰ -12 00	DR LAURA F. MAREK Sunflower Genetic Resources	Determination of Textural, Rheological Properties and SFC, SMP Values of Oleogels Prepared Using Sunflower Oil – H. PEHLİVANOGLU, O. S. TOKER, H. IMAMOGLU, M DEMIRCI	Effect of different storage conditions on quality properties of raw and roasted sunflower kernels – F. SEYHAN, M. B. BAHAR, B. TOPAL, B. ÖZTÜRK, F. S. BAYRAKTAR
1200-12 20	MULPURI	Assessment of sunflower oil adulteration – A. CEVIK, A. UNVER	The Evaluation of Sunflower Harvest Waste as Silage Feed – S. BUYUKKILIC BEYZI, M. YILMAZ, Y. KONCA
12^{20} - 12^{30}	Discussion	Discussion	Discussion
12^{30} -13 30	Lunch (Courtesy of Edirne Commodity Excha	inge)	
13 ³⁰ -15 ³⁰	9 th Session Chair: DR ABELARDO DE LA VEGA	9 th Session Chair: PROF DR NURHAN T. DUNFORD	9 th Session Chair: PROF DR SEVGI CALISKAN
13 ³⁰ -13 ⁵⁰	Invited Speaker DR NADA HLADNI	The effects of vacuum and atmospheric deep-fat frying process on total frying-use time of sunflower oil and on french fries quality – E. DEVSEREN, D. TOMRUK, U. BAYSAN, M. KOC, H. KARATAŞ, F. ERTEKIN	Study of the characteristics of cultivated varieties of sunflower, regarding the production of high quality sunflower meal with dehulling process - S. DAUGUET, F. LABALETTE, F. FINE, P. CARRE, A.MERRIEN, J. P. PALLEAU
13 ⁵⁰ -14 ¹⁰	Present status and future prospects of global confectionery sunflower production	Effect of curcumin nanoparticles on oxidative stability of sunflower oil-in-water emulsions – F. BOZKURT, M. T. YILMAZ, C. YILDIRIM	Acceptability of chapati Made With Supplementation of Sunflower (<i>Helianthus annuus</i> L.) Seed Meal – M. KARWASRA, S. DHIYA

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14 ¹⁰ -14 ³⁰	Grain, kernel and hull characterization of oilseed and oilseed x confectionary genotypes-S. ZUIL, M. LAUREANO, P. ROCCA, M. DELLA MADDALENA	Application of artificial neural network on prediction of moisture content of the deep-fat frying of beef meatballs in sunflower oil-H.I. KOZAN, C. SARIÇOBAN, H. AKYÜREK	Some Antinutrients and in vitro Protein Digestibility of Home Processed Sunflower Seed Meal – M. KARWASRA, S. DHIYA
14 ³⁰ -14 ⁵⁰	Effects of herbicide and salinity stresses on some defense responses of sunflower plant- A. KAYA	Effect of the Deep-Fat Frying Process on Aroma Compounds of Sunflower Seed Oil – S. KESEN, A. S. SÖNMEZDAĞ, A. AMANPOUR, H KELEBEK, S. SELLI	
14^{50} -5 00	Discussion	Discussion	Discussion
15^{00} - 15^{30}	Coffee break	Coffee break	Coffee break
15^{30} - 17^{00}	10 th Session Chair: DR PIERRE CASADEBEIG	10 th Session Chair: DR SUSAN THOMPSON	10 th Session Chair: DR NICOLAS LANGLADE
15 ³⁰ -15 ⁵⁰	Quantitative Determination of Sunflower in Mixed Concentrate Feeds by Real Time PCR-M. KAYA,Z. KIYMA	The Effect of the ESSENTIAL OIL from Citrus aurantium as a source of natural antioxidant in sunflower oil – O. ERDOĞDU, A. BOZDOGAN	
15 ⁵⁰ -16 ¹⁰	The evaluation of annual wild <i>Helianthus</i> species for their morphological, phenological and seed chemical characteristics in field conditions – F. ONEMLI, G. ONEMLI	LC-DAD/ESI-MS/MS Characterization of Phenolic Compounds of Sunflower oil – H. KELEBEK, S. SELLI, A. S. SÖNMEZDAĞ, S. KESEN, G. GUCLU, O. KOLA	The Meeting of International Consortium for Sunflower Genomic Resources
16 ¹⁰ -16 ³⁰		Lessons from ten years of an interprofessional survey plan on sunflower food safety - S. DAUGUET, F. LACOSTE	
16^{30} - 16^{45}	Discussion	Discussion	

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16 ⁴⁵ -17 ⁴⁵	ISA GENERAL ASSEMBLY					
17 ⁴⁵ -18 ⁰⁰	Closing Ceremony					
19 ³⁰ -23 ³⁰	GALA DINNER					

	02.06.2016 THURSDAY					
09 ³⁰ -12 ⁰⁰	Field Day in Trakya Agricultural Research Institute Visiting Demo Plots					
1200-1300	Lunch					
13 ³⁰ -17 ³⁰ Edirne City Tour						
17 ³⁰ -	Free Shopping Time					

03.06.2016 FRIDAY			
07^{00} - 19^{30}	Istanbul City Tour		
19 ³⁰ -23 ³⁰ Bosphorus Yacht Tour and Dinner			

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EFFICACY OF TRICHODERMA SPP. ISOLATES AGAINST SCLEROTINIA SCLEROTIORUM ON SUNFLOWER SEEDLINGS

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ABSTRACT

Trichoderma species are well known as effective antagonists to a variety of soil fungal pathogens. The aim of this research was to test the ability of *Trichoderma* spp. isolates, which previously indicated antagonistic activity (Tančić et al. 2012), to protect sunflower seedlings from Sclerotinia sclerotiorum. Ten Trichoderma spp. isolates obtained from different soil types and localities in Vojvodina province and one S. sclerotiorum isolate from sunflower grown at Rimski Šančevi (Serbia) were used in research. Biological efficacy was tested on 100 sunflower seeds treated with *Trichoderma* spp. suspensions (1x10⁶) in two different treatments: T-30 (modified Mukhtar et al., 2012) and T-1.2 (Maslienko, 2005). Trichodermacoated seeds were placed in four replicates on wet filter paper in Petri dishes. Next to each Trichoderma-coated seed the 5 mm² plug of S. sclerotiorum mycelia was placed, and incubated under the optimal laboratory conditions. Seeds treated with sterile distilled water with pathogen and without it were used as a positive and negative control, respectively. After seven days, biological efficacy of Trichoderma spp. isolates was assessed and calculated according to Liu et al. (2009). According to obtained results, biological efficacy of all tested Trichoderma isolates was statistically significant as compared to the positive control in both treatments. Good antagonism with over 50% of biological efficacy was registered in 8 isolates in T-30, and 3 isolates in treatment T-1.2. Three Trichoderma isolates which showed biological efficacy over 50% in both treatments can be considered as potential biocontrol agents which should be included in further more comprehensive research.

Key words: Trichoderma, Sclerotinia sclerotiorum, Antagonism, Sunflower

INTRODUCTION

Sunflower (*Helianthus annuus* L.) is one of the most important annual species grown in Serbia mostly for its edible oil. Areas grown under sunflower were around 170 000 hectares with the expected yield of 422 000 tones and sunflower oil production of 139 000 tones in economical year 2015/2016 (Chamber of Commerce and Industry of Serbia, 2016).

Farmers around the world are familiar with *S. sclerotiorum* (Lib.) de Bary as a threat to numerous crops such as sunflower, soybean, oilseed rape, edible dry bean, chickpea, peanut, dry pea, lentils and various vegetables as well. Occurrence of diseases caused by *S. sclerotiorum* on sunflower is influenced by genotype and weather conditions. Moist and cold weather conditions prevailing in temperate climate regions favours *S. sclerotiorum* development. In Serbia, weather conditions favoured economically important Sclerotinia development on sunflower head in 1999 and 2005 with diseased plants even over 60% (Maširević and Forgić, 2000; Maširević and Dedić, 2006). Diseases caused by this fungus can

appear during the whole sunflower growing season, and yield loss depends on the sunflower development stage in which the disease occurs. Sunflower plants infected at the beginning of flowering stage can lose up to 98% of their potential yield, while plants infected eight weeks after flowering can lose not more than 12% of their potential yield (Maširević & Gulya 1992). The major control method for *Sclerotinia* diseases has been fungicide application in combination with host resistance.

Considering growing demand for organic food production, biocontrol of such cosmopolitan and devastating pathogen is a big challenge. The use of biofertilizers and biopesticides is an alternative for sustaining high eco-friendly production. Integrated control is facilitated by the fact that *Trichoderma* species are resistant to most chemical pesticides (Harman, 2011; FRAC, 2016). Trichoderma species have been known since 1930s (Weindling, 1932), but since 1990s their usage in commercial agriculture has been increased (Harman 2004, 2011). So far, Trichoderma species have been known as effective on nutrient utilization with high reproductive potential which allow them to survive under unfavourable conditions and makes them very competitive. Presence of great variety of lytic enzymes (cellwall degrading enzymes) and secondary metabolites (gliotoxin, gliovirin, viridin, viridiol etc.) makes Trichoderma strongly aggressive to broad range of phytopathogenic fungi (Vinale et al., 2008). The main biocontrol mechanisms of Trichoderma species, when direct confrontation with pathogen occurs, are mycoparasitism and antibiosis (Howel, 2003). Another mechanism which is quite effective as well, but do not consider direct confrontation with the pathogen, is competition for soil nutrients and space. Additionally, Trichoderma species are known as well as plant growth promoter agents and promoters of plant defense mechanisms (Shoresh et al., 2010; Harman, 2011). Trichoderma colonizes roots and provides at least season-long benefits to plants, although it can be even for life because the best strains fully colonize roots as they grow (Harman 2000). So far it is recorded that Trichoderma species improve growth of lettuce, tomato, pepper, wheat, maize, soybean, chilli (Vinale et al., 2004; Tucci et al., 2011; Sukla et al., 2015; Maisuria and Patel, 2009; Asaduzzaman et al., 2010). Also, Trichoderma spp. is stimulating defense responses in its host plants and is known as one of the best induced systemic resistance (ISR) agents (Shoresh et al., 2010; Shoresh, 2005).

Trichoderma species are mainly soil fungi found in agricultural soils, native prairie, forests, salt marsh, desert soils of all climatic zones, but also in dead plant material, living roots of various plant species, seeds, lake water and air (Monte, 2001). World-wide distribution, fast growth and high spore production make those species easy to find and isolate. After all, one should bear in mind that not all Trichoderma strains are effective, most of them are not, and some may even be phytotoxic or pathogenic (Menzies, 1993), so strain selection is of crucial importance. Given that, the aim of this study was to test ability of native Serbian Trichoderma strains to protect sunflower seedlings in early stage from pathogen S. sclerotiorum.

MATERIAL AND METHODS

Plant and fungal material used: Fungal material was obtained from soil samples originated from different soil types and localities in Serbia, mainly from Vojvodina province. All *Trichoderma* spp. isolates were refined to single-spore according to Leslie and Summerell (2006). Ten isolates which previously indicated good antagonistic activity in dual culture test (Tančić et al., 2012) were selected for this research. Pathogen *S. sclerotiorum* was isolated from diseased sunflower plant at Rimski Šančevi.

Trichoderma isolates' efficacy against *S. sclerotiorum* was tested on sunflower seeds of sterile parental line VL-A-8A.

Preparation of conidia suspension: A conidia suspensions of ten tested *Trichoderma* isolates were prepared from 7-days old isolates by flooding method. Such suspensions were filtered through cheesecloth, and conidial concentrations were adjusted to 10^6 conidia/ml by Neubauer's haemocytometer. Additionally, suspensions were amended according to Mukhtar et al. (2012) method.

Treatment T-1.2 considered that seeds were treated with 1.2 μ l of *Trichoderma* suspension which was equally distributed per g of seeds and air dried on filter paper in Petri plates for 24 hours at room temperature (Маслиенко, 2005). Control was treated with 1.2 μ l of sterile distilled water per g of seeds.

Treatment T-30 considered that seeds were dipped in seed-coating suspensions for 30 minutes and air dried on filter paper in Petri plates for 24 hours at room temperature, while sterile distilled water was used as a control.

Biological efficacy test: was done on 100 sunflower seeds treated with *Trichoderma* suspensions of different intensities (T-1.2 and T-30). Treated seeds were germinated in four replicates on double wet filter paper. Next to each sunflower seed, the 5 mm² plug of potato dextrose agar (PDA) with 7-day old micelia of *S. sclerotiorum*, was placed. Seeds treated with sterile distilled water without presence of pathogen *S. sclerotiorum* plugs were used as negative control, while seeds treated with sterile distilled water with presence of *S. sclerotiorum* plugs were used as positive control. Seeds were germinated in growth chamber with 12h photoperiod at $25\pm1^{\circ}$ C. After seven days diseased seedlings and seeds were counted, and biological efficacy of the *Trichoderma* isolate was calculated according to formula (Liu et al. 2009):

$$C(\%) = 100 * (a - b) / a$$

where C is biological efficacy in %, a – number of diseased seeds and seedlings in positive control, and b – number of diseased seeds and seedlings in treatment.

Beside biological efficacy, germination (G) was calculated as well on 7th day of the experiment.

Statistical analyses: All obtained data were analyzed in Statistica 12 using Duncan's test (percentages were previously transformed in $ArcSin\sqrt{\%}$).

RESULTS AND DISCUSSION

Formation of rhizosphere microflora occurs usually in first three days after germination, and its progress in the deeper soil layers follows root growing and stimulates plant exometabolites at the same time (Acatypoba, 2009). This is very important in biocontrol especially because young seedlings are often infected by pathogens in early stage of their development. Due to above mentioned, biological efficacy was estimated in the first days of sunflower germination and expressed as a percentage of protected seeds and seedlings comparing positive control (seeds without *Trichoderma* treatment grown in presence of pathogen *S. sclerotiorum*).

Germination was calculated on 7^{th} day of incubation. Lower germination rates were registered in treatments with lower biological efficacy. Biological efficacy of all tested isolates was statistically significant as compared to the positive control in both treatments. According to obtained results, biological efficacy of tested *Trichoderma* isolates varied from 36-68% and 23.8-60.6% for treatments T-30 and T-1.2 respectively (Table 1). Excellent

antagonism with over 50% of biological efficacy was registered in 8 isolates in T-30, and 3 isolates in treatment T-1.2 (bold values in Table 1). Three *Trichoderma* isolates – K150, K173 and K174 showed biological efficacy over 50% in both treatments. These are promising results considering that some authors with bacterial antagonist reached biological efficacy against *Fusarium* spp. on sunflower seedlings from 0-36% (Acatypoba, 2009), while biological efficacy of fungal and bacterial antagonist against *S. sclerotiorum* on sunflower stem under the field conditions was much higher - 54.5-100% (Φμρcob et al., 2009). Besides on sunflower, the antagonistic activity of *Trichoderma* spp. against *S. sclerotiorum* was proven on other crops as well. Thus, the application of *T. harzianum* as alignate capsules increased the survival of soybean plants more than 100% and 40% in greenhouse and in the field, respectively (Menendez and Godeas, 1998). Isolates of *T. harzianum* also protected over 80% of tomato, squash and eggplant seedlings inoculated with *S. sclerotiorum* in the greenhouse experiments (Abdullah et al., 2008). Further, *T. virens* significantly reduces the percentage of viable sclerotia and number of apothecia produced (Huang and Erickson, 2008) which can be used for bioregulation of pathogen density in soil.

Table 1. Biological efficacy of two different treatments with *Trichoderma* spp. isolates against *S. sclerotiorum* on sunflower seedlings

	T-30		T-1.2	
Isolate No	C (%)	G (%)	C (%)	G (%)
K114	50.0ab	86 ^{ab}	26.2 ^{bc}	78ª
K132	36.0 ^b	74 ^a	45.2ª	84 ^{ab}
K150	58.0ac	94 ^{ab}	58.3 ^a	80 ^{ab}
K160	50.0 ^{ab}	90 ^{ab}	40.5 ^{abc}	77ª
K173	56.0 ^{ac}	76 ^a	54.8a	85 ^{ab}
K174	58.0 ^{ac}	86 ^{ab}	60.6 ^a	85 ^{ab}
K175	68.0°	74 ^a	23.8 ^b	91 ^b
K176	59.5 ^a	88 ^{ab}	48.0 ^{ab}	87 ^{ab}
K178	40.0 ^b	90 ^{ab}	42.9 ^{ac}	80 ^{ab}
K179	53.6 ^a	98 ^b	44.0 ^{ab}	87 ^{ab}
- Control	100 ^d	88 ^{ab}	100 ^d	96 ^{ab}
+ Control	$0.00^{\rm e}$	80^{ab}	0.00^{e}	82 ^{ab}

Legend: Values in the columns followed by the same letters are not significantly different (p<0.05) by Duncan's test; Values are average of four replicates;

Beside S. sclerotiorum, it has been proven that Trichoderma spp. are aggressive to broad range of phytopathogenic fungi – Rhizoctonia, Fusarium, Alternaria, Ustilago, Venturia, Colletotrichum, Pythium, Phytophthora, Thielaviopsis, Sclerotium cepivorum, Sclerotinia minor etc. (Vinale et al., 2008; Thomas et al., 2004; McLean et al., 2012).

All mentioned above is leading to a conclusion that those perspective isolates from our research could also be good antagonists for some other important sunflower pathogens which should be tested in some further research.

CONCLUSION

Three out of ten tested *Trichoderma* isolates originating from Serbia expressed excellent ability to protect sunflower seedlings from pathogen *S. sclerotiorum* in both treatments. Those isolates can be considered as potential biocontrol agents and should be included in further, more comprehensive, research.

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